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Delivering on the therapeutic potential of CRISPR/Cas9: Development of an LNPmediated genome editing therapeutic for the treatment of ATTR

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October 18, 2018

26th Annual Congress of the European Society of Gene and Cell Therapy



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This press release contains "forward-looking statements" of Intellia Therapeutics, Inc. ("Intellia") within the meaning of the Private Securities Litigation Reform Act of 1995. These forward-looking statements include, but are not limited to, express or implied statements regarding Intellia's ability to advance and expand the CRISPR/Cas9 technology to develop into human therapeutic products, as well as our CRISPR/Cas9 intellectual property portfolio; our ability to achieve stable or effective genome editing; our ability to administer multiple doses of our CRISPR/Cas9 product candidates; the potential timing and advancement of our preclinical studies, including continuing non-human primate studies for our Transthyretin Amyloidosis ("ATTR") program and other programs (such as alpha-1 antitrypsin deficiency (AATD)), and clinical trials; the timing and potential achievement of milestones to advance our pipeline; our ability to replicate results achieved in our preclinical studies, including those in our ATTR, AATD and Wilms' Tumor 1 (WT1) programs, in any future studies, including human clinical trials; the potential development of other in vivo or ex vivo cell therapeutics of all types, and those targeting WT1 in particular, using CRISPR/Cas9 technology; our ability to continue to conduct successful Investigational New Drug ("IND") enabling studies of a lead ATTR development candidate and subsequently submitting an IND application by the end of 2019 that will be accepted by the regulatory agencies; our intent to present additional data for organs beyond the liver, additional insertion/repair data, and preclinical data in support of our first ex vivo programs on immuno-oncology and autoimmune/inflammation indications during 2018; the expansion of our fully automated bioinformatics platform; our ability to advance a development candidate for a second indication by late 2018; our potential ability to conduct a pre-IND meeting with the U.S. Food and Drug Administration ("FDA") for ATTR; the intellectual property position and strategy of Intellia's licensors or other parties from which it derives rights; actions by government agencies; the impact of our collaborations on our development programs; the potential timing of regulatory filings regarding our development programs; the

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Hereditary Transthyretin Amyloidosis (hATTR) Is a Rare Disease Typically Fatal if Untreated





About ATTR

- Autosomal dominant disease²
- Caused by misfolded transthyretin (transports thyroxine and retinol-binding protein), which affects nerves, heart, kidneys and eyes
- >100 known mutations with V30M and V122I among the most common associated with the diseases^{1, 3}
- Estimated 50,000 hATTR patients worldwide¹
- Typically fatal within 2-15 years from onset of symptoms¹

¹Ann Med. 2015;47(8):625-38. ²Annu Rev Med. 2000; 51:543-569. ³Handbook of Clinical Neurology. 2013:115(38).



Intellia's Modular Lipid Nanoparticle (LNP) System Delivers CRISPR/Cas9 to Make an *In Vivo* Edit





Intellia's Modular Lipid Nanoparticle (LNP) System Delivers CRISPR/Cas9 to Make an *In Vivo* Edit

Intellia's LNP delivery system includes a single guide RNA, mRNA encoding *S. py.* Cas9 and a lipid formulation encapsulating these





Editing *in vivo* requires cargo release, mRNA translation, RNP assembly and Cas9 import into the cell's nucleus



Durable Liver Editing and Knockdown of TTR Persists 12 Months in Mice with No Histological Findings



Single administration given at Day 1





No transformation or neoplastic changes observed across >100 mice over time up to 12 months post-edit



Liver Editing Efficiency Is Not Affected by Pre-Dosing with LNP-mRNA Cas9



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Mouse Multi-Dose Study: Repeat Low Dose of LNPs with CRISPR/Cas9 Is Comparable to Single High Dose



Multiple weekly or monthly doses of 0.5 mg/kg are comparable to one 2 mg/kg dose



Editing Strategy Relies on Knockout Caused by Error-Prone Non-Homologous End Joining (NHEJ) of a Double-Strand Break in Liver Cells



CRISPR Double-Stranded Break

NHEJ Repair with INDELS



Lead Human TTR CRISPR/Cas9 LNPs Demonstrate On-Target Editing, and Reduction of mRNA and TTR Protein in Primary Human Hepatocytes *In Vitro*





Top Human Guides Exhibit Robust, Dose-Responsive Liver Editing and Reduction of TTR in huTTR Mice



VelociGene[®] is a registered trademark of Regeneron Pharmaceuticals, Inc.



Findings from huTTR V30M Mouse Model Study Recapitulate TTR Deposition Phenotype in Tissues and the Nervous System



Mouse model from Santos et al. 2003; In collaboration with U. of Porto



Decreasing Serum TTR by Editing the huTTR V30M Mouse Model Via CRISPR/Cas9 LNP Dramatically Decreases Amyloid Deposition in Tissues



Mouse model from Santos et al. 2003; In collaboration with U. of Porto



Therapeutically Relevant Reduction of Serum TTR Protein Achieved in Initial Non-Human Primate (NHP) Studies After a Single Dose of LNPs with CRISPR/Cas9



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LNPs and Cargo Exhibit 18-24 Hour $T_{1/2}$ and Are Cleared from Circulation and Liver Within 5 days in NHP



High Correlation Achieved Between Liver Edit and Reduction of TTR; >35-40% Liver Edit Needed to Achieve Therapeutically Meaningful Reduction of TTR



Fragments taken from 4
lobes of liver





Summary

- LNPs encapsulating CRISPR/Cas9 components targeting human TTR enable significant editing of the TTR gene across multiple species, including mice and NHPs
- Following a single dose of LNP-delivered CRISPR/Cas9 in mice:
 - Editing levels achieved that resulted in >97% reduction in circulating serum TTR protein
 - Reduction of circulating levels of TTR sustained for at least 12 months
 - No significant histopathology findings noted
- Humanized mouse model of hATTR that expresses the V30M mutant form of the human TTR protein demonstrated rescue of amyloid deposition in multiple tissues after a single dose of LNPs containing the CRISPR/Cas9 components
- In NHPs, achieved a therapeutically meaningful level of TTR protein reduction that correlated with robust and significant editing in the liver
- S. py. Cas9 mRNA, sgRNA and ionizable lipid are quickly cleared from circulation, with the lipid having plasma and liver half-lives of 20 hours and 17 hours, respectively, in NHPs
- Demonstrated the potential of LNP delivered in vivo CRISPR/Cas9 gene editing; suggests that future therapies based on this platform may enable next-generation, curative treatment paradigms for chronic genetic diseases such as ATTR



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