CRISPR/Cas9-mediated Gene Knockout to Address Primary Hyperoxaluria

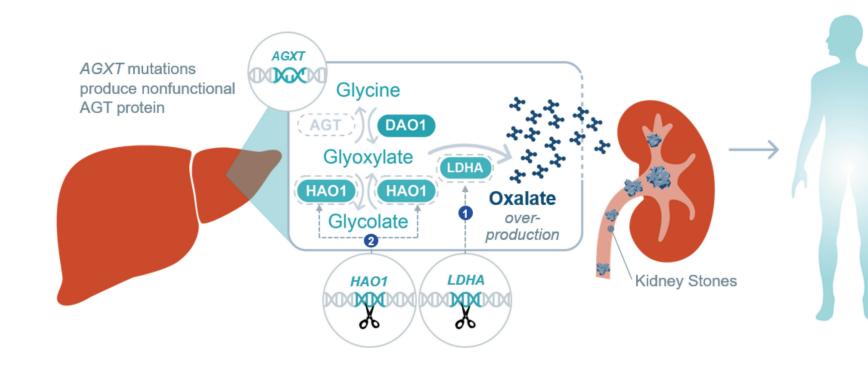
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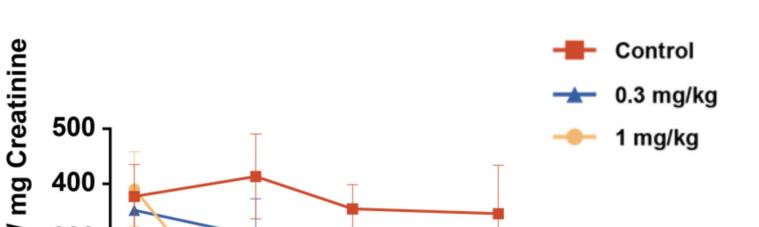
INTRODUCTION

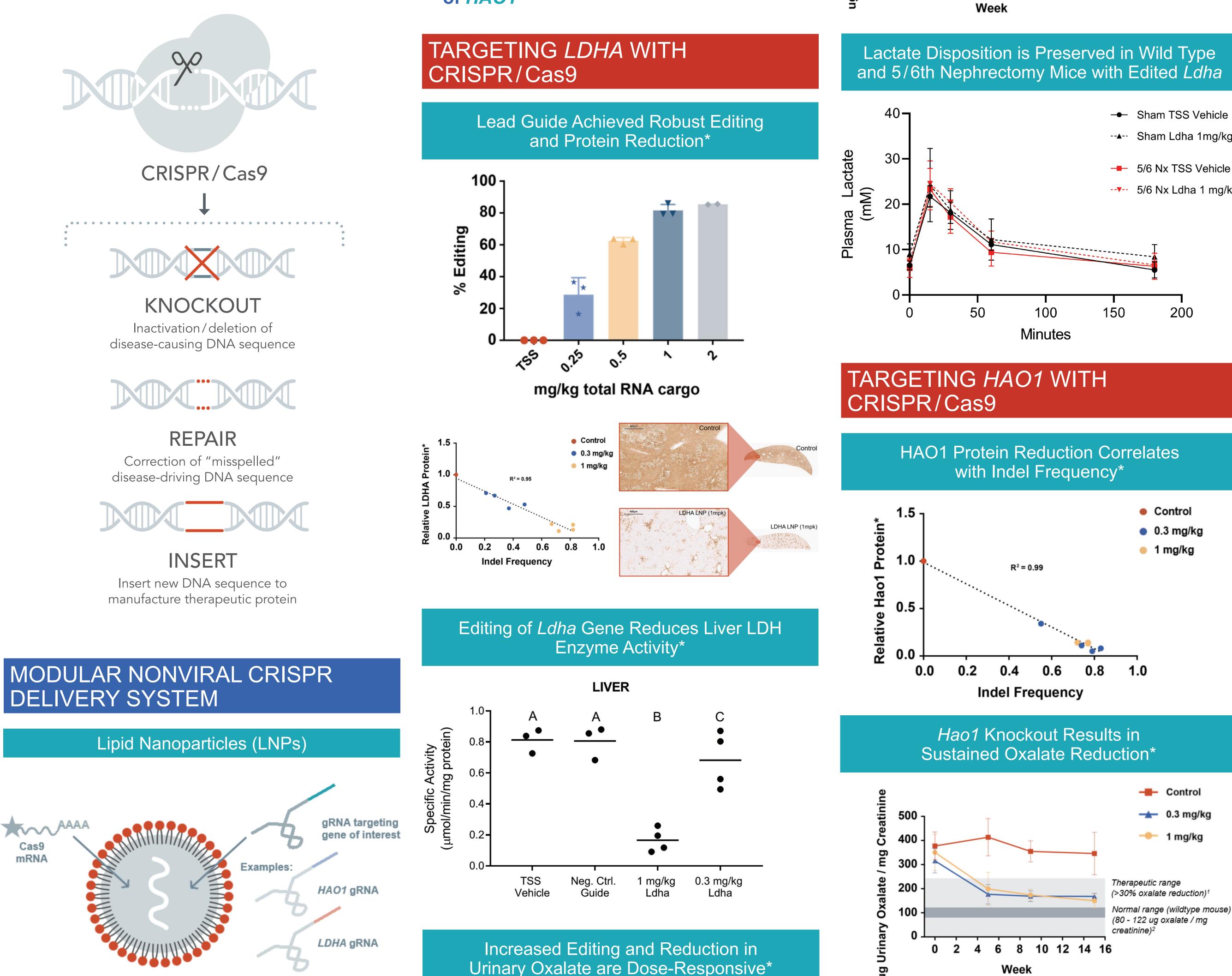
Primary hyperoxaluria (PH) is a rare genetic disease caused by mutations in one of three genes (AGXT, GRHPR and HOGA1) involved in the glyoxylate detoxification pathway, giving rise to PH types 1, 2, and 3, respectively. PH is characterized by excessive accumulation of the toxic waste product oxalate, which leads to formation of insoluble deposits in the kidney and other organs, resulting in renal failure and systemic oxalosis. Currently, the treatment of late-stage disease is limited to combined liver-kidney transplantation. Here, we tested the hypothesis that non-viral CRISPR/Cas9mediated editing of two genes involved in oxalate formation, Ldha and Hao1, could significantly lower urinary oxalate in a mouse model of PH1, providing proof-of-concept for a one-time treatment approach for the disease.

RESULTS



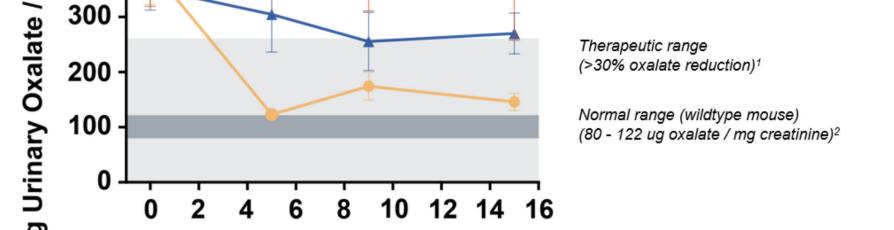
Ldha Knockout Results in Sustained Oxalate Reduction*



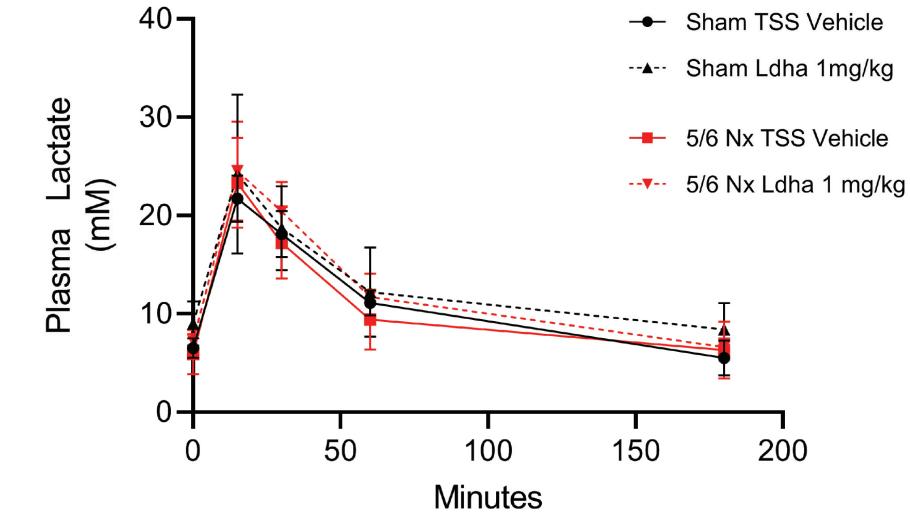


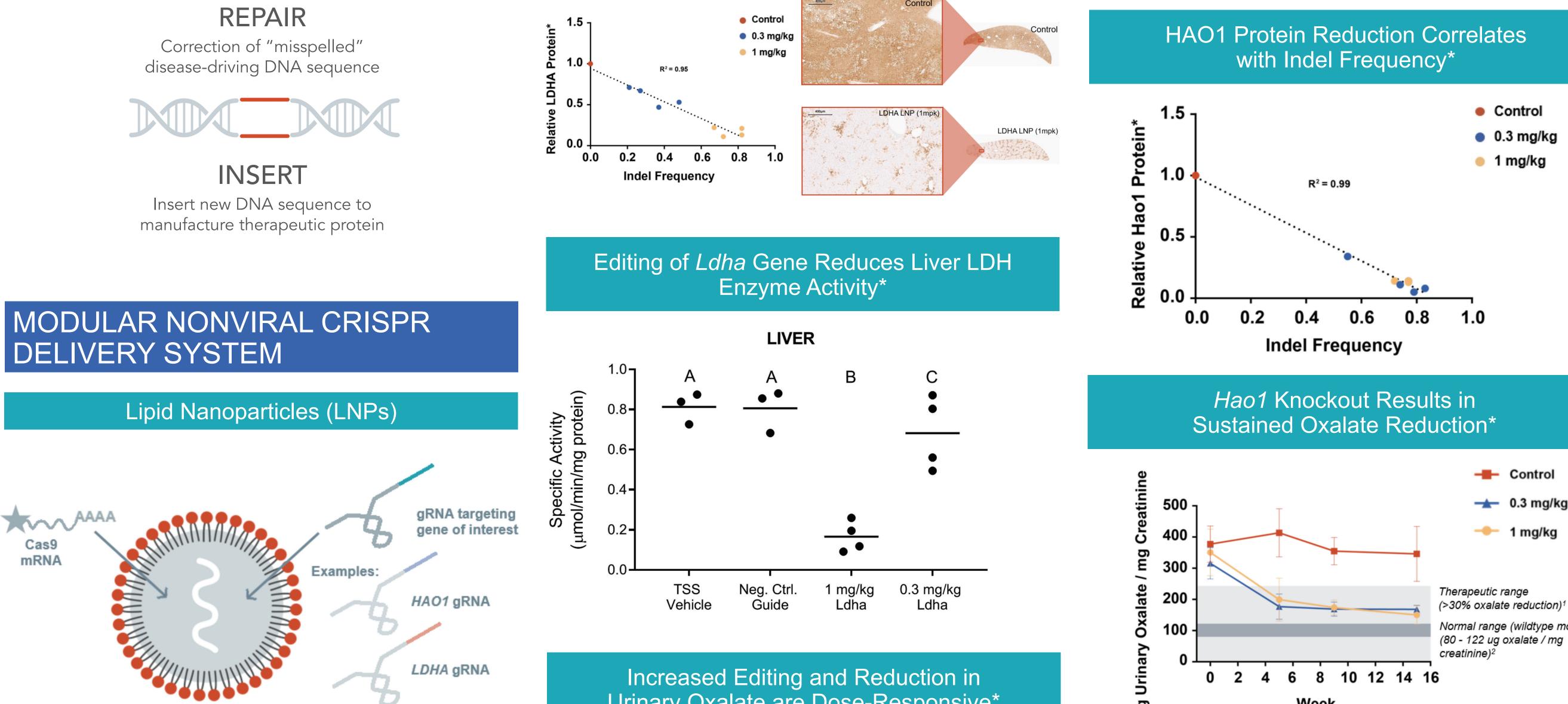
Potential to treat PH1 with either: 1. CRISPR/Cas9-mediated knockout

- of LDHA or
- 2. CRISPR/Cas9-mediated knockout of HAO1



Lactate Disposition is Preserved in Wild Type and 5/6th Nephrectomy Mice with Edited Ldha

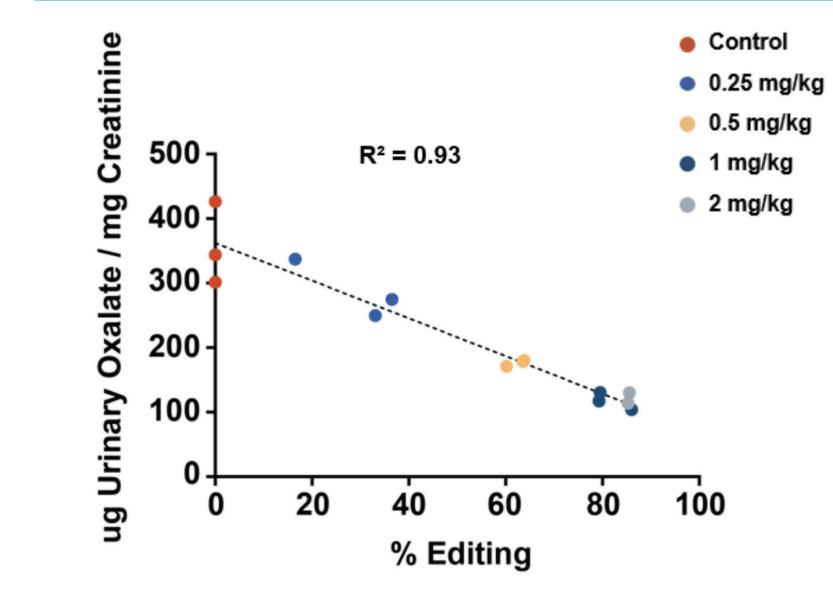




Variable portion of Intellia's modular **LNP-based liver knockout approach** is limited to 20-mer of gRNA

Key Advantages of LNP Delivery

- Large cargo capacity for CRISPR/Cas9
- Transient expression
- Low immunogenicity with redosing capability
- Biodegradable and well-tolerated
- Scalable synthetic manufacturing



*Agxt^{-/-} mouse

CONCLUSIONS

- Modular LNP-based CRISPR system enables efficient knockout of genes involved in oxalate production
- Single treatment targeting either *Hao1* or *Ldha* leads to a dose-dependent and persistent reduction of urinary oxalate levels in the *Agxt*^{-/-} mouse model of PH1
- Ldha gene disruption decreased LDH enzyme activity in the liver, yet did not impair the disposition of lactate in either wild type or renally-impaired mice

• These results suggest the promise of LNP-delivered CRISPR for treating genetic forms of hyperoxaluria using a single-course treatment paradigm





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